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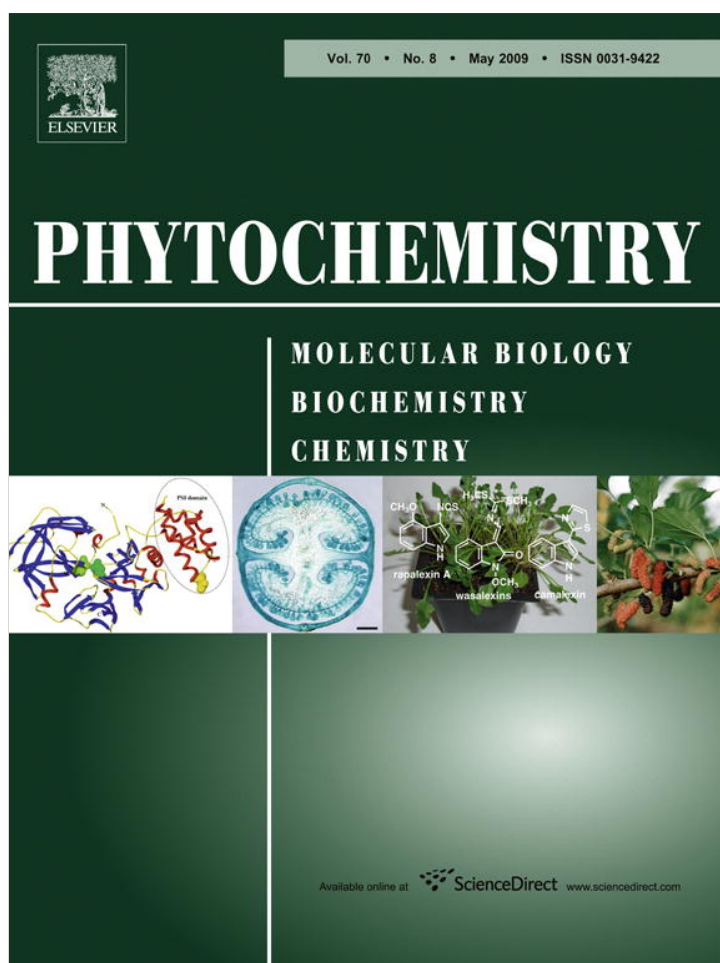
Published in:
Phytochemistry

DOI:
[10.1016/j.phytochem.2009.05.004](https://doi.org/10.1016/j.phytochem.2009.05.004)

Publication date:
2009

Document version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Stærk, D., Kesting, J. R., Sairafianpour, M., Witt, M., Asili, J., Emami, S. A., & Jaroszewski, J. W. (2009). Accelerated dereplication of crude extracts using HPLC-PDA-MS-SPE-NMR. *Phytochemistry*, 70(8), 1055-1061. <https://doi.org/10.1016/j.phytochem.2009.05.004>



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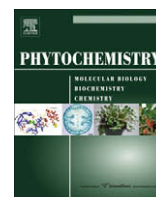
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Phytochemistry

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Accelerated dereplication of crude extracts using HPLC–PDA–MS–SPE–NMR: Quinolinone alkaloids of *Haplophyllum acutifolium*

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ARTICLE INFO

Article history:

Received 9 March 2009

Received in revised form 15 April 2009

Available online 18 June 2009

Keywords:

Haplophyllum acutifolium

Haplophyllum perforatum

Rutaceae

HPLC–PDA–MS–SPE–NMR

Haplacutines A–F

NMR

Dereplication

ABSTRACT

Direct hyphenation of analytical-scale high-performance liquid chromatography, photo-diode array detection, mass spectrometry, solid-phase extraction and nuclear magnetic resonance spectroscopy (HPLC–PDA–MS–SPE–NMR) has been used for accelerated dereplication of crude extract of *Haplophyllum acutifolium* (syn. *Haplophyllum perforatum*). This technique allowed fast on-line identification of six quinolinone alkaloids, named haplacutine A–F, as well as of acutine, haplamine, eudesmine, and 2-nonylquinolin-4(1H)-one. Acutine and haplacutine E, isolated by preparative-scale HPLC, showed moderate antiparasmodial activity with IC₅₀ values of $2.17 \pm 0.22 \mu\text{M}$ and $3.79 \pm 0.24 \mu\text{M}$, respectively (chloroquine-sensitive *Plasmodium falciparum* 3D7 strain).

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1. Introduction

Selecting the strategy for isolation of constituents from plant extracts and other natural sources is of utmost importance with respect to the outcome as well as time and efforts spent on discovery of pharmacologically active natural products. Prior to preparative-scale isolation, it is convenient to gain as much knowledge as possible about chemical entities present, both in order to make qualified decisions about worthwhileness of isolation of a particular constituent, and to choose optimal isolation methodology. Although some information about extract constituents can be obtained from thin layer chromatography combined with the use of compound class-specific spray reagents, by gas chromatography combined with mass spectrometry (MS), and by high-performance liquid chromatography (HPLC) combined with photo-diode array detection (PDA) or MS, all these techniques suffer from the limited structural information they provide as compared to nuclear magnetic resonance (NMR) spectroscopic techniques.

Within recent years, hyphenation of HPLC, PDA, MS, and solid-phase extraction (SPE), with NMR spectroscopy (HPLC–PDA–MS–

SPE–NMR) has emerged as an attractive analytical technique for fast and reliable dereplication of crude plant extracts (Jaroszewski, 2005a,b; Staerk et al., 2006; Sturm and Seger, 2007). This technique has been used for identification of known compounds as well as for rigorous structure elucidation of new natural products directly from crude extracts. Examples include sesquiterpene lactones (Lambert et al., 2007), quinic acid derivatives (Sprogø et al., 2007), diterpenes and caffeoyl glycosides (Clarkson et al., 2006a, 2006b), flavonoid glycosides (Clarkson et al., 2005; Exarchou et al., 2006; Tatsis et al., 2007), cardenolide glycosides (Clarkson et al., 2005), isoflavonoids (Lambert et al., 2005a), and alkaloids (Bieri et al., 2006; Lee et al., 2007; Sturm et al., 2007). One advantage of the HPLC–SPE–NMR technique compared to direct HPLC–NMR methods (Albert, 2002; Jaroszewski, 2005a) is the possibility of linear analyte accumulation by repetitive adsorption on SPE cartridges (Clarkson et al., 2006a; Lambert et al., 2005a, b). Subsequent elution of the analyte with a deuterated organic solvent, resulting in focusing of the analyte in the detection cell, adds further increase in sensitivity of this hyphenated NMR technique (Jaroszewski, 2005b; Staerk et al., 2006). Here, we describe application of HPLC–PDA–MS–SPE–NMR to fast dereplication of an extract of aerial parts of *Haplophyllum acutifolium*, which showed activity (IC₅₀ < 12 $\mu\text{g/ml}$) in our *in vitro* *Plasmodium falciparum* toxicity screening assay.

Haplophyllum A. Juss. (Rutaceae) is a genus of small flowering perennial plants comprising approximately 50 species, distributed

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from the Mediterranean parts of Europe and Africa to Eastern parts of Siberia. Around 30 of these species are represented in Iran, 14 of which are endemic to the country. *Haplophyllum* is known as a rich source of quinoline alkaloids, and several phytochemical investigations of different species have been reported in recent years (Michael, 2008), including investigations of *H. acutifolium* collected in Turkmenistan (Razzakova et al., 1975, 1986, and references cited therein). Extracts of various *Haplophyllum* species are used in traditional medicine as analgesics, antispasmodics, diuretics, and sedatives, as well as topical agents against skin diseases (Nazrullaev et al., 2001). Quinolinone alkaloids isolated from *Haplophyllum* and other Rutaceous species have shown estrogenic (Nazrullaev et al., 2001), antifungal (Cantrell et al., 2005), antialgal (Cheng et al., 2005), antiviral (Cheng et al., 2005), antibacterial (Hanawa et al., 2004), and antiparasitic activity (Michael, 2003). In spite of the existence of an abundant literature about quinolinone alkaloids of *Haplophyllum*, dereplication of the extract of *H. acutifolium* using HPLC–PDA–MS–SPE–NMR allowed fast identification of several previously unknown minor alkaloids present in the plant.

2. Results and discussion

HPLC–PDA–MS–SPE–NMR experiments with crude ethyl acetate extract of aerial parts of *H. acutifolium* (crude extract A, see Section 4) were performed using UV trace at 235 nm for threshold-based adsorption of constituents on SPE cartridges containing poly(divinylbenzene)-based resin (Clarkson et al., 2007). Four or six repetitive trappings of material eluted in peaks 1–9 (Fig. 1) were performed. The trapped compounds were subsequently eluted with CD₃CN for acquisition of NMR data. This led to identification of five new and two already known 4-quinolinone alkaloids (**1**, **2a**, **3**, **4**, **6**, **8**, and **9**) along with a tricyclic hemiterpenoid alkaloid haplamine (**5**) and the lignan **7** as detailed below; the identified structures (Fig. 2) are numbered according to elution order of the HPLC peaks (Fig. 1). Parallel HPLC–PDA–MS–SPE–NMR experiments with an alkaloid-enriched fraction (fraction III from crude extract B, see Section 4) showed the existence of one additional, new quinolinone alkaloid (**2b**) as a minor constituent.

The constituent eluted as peak 1 (compound **1**) was assigned the molecular formula C₁₈H₂₁NO₂ as determined by off-line HRESIMS. The ¹H NMR spectrum acquired in the HPLC–SPE–NMR mode showed the presence of a 1,2-disubstituted benzene with a characteristic downfield shift of H-5 (δ 8.11, J₅₋₆ = 8.0 Hz and J₅₋₇ = 1.4 Hz) attributable to a *peri*-oriented carbonyl group, a one-proton singlet of H-3 (δ 5.98), and a broad singlet of H-1 (δ 9.55). This information, together with ¹H–¹H correlations for H-5, H-6, H-7, and H-8 found in a COSY spectrum and ¹³C NMR reso-

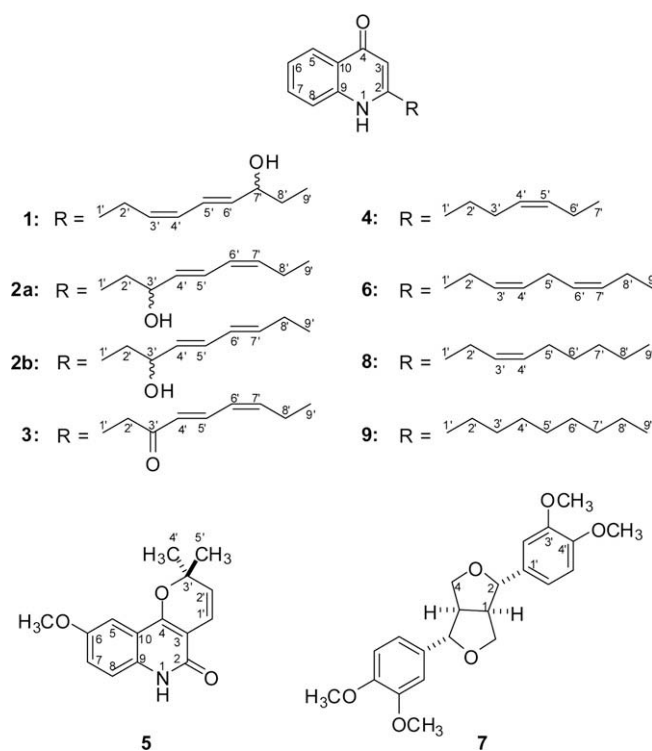


Fig. 2. Structures of compounds 1–9.

nances identified in a HSQC spectrum, both acquired in the HPLC–SPE–NMR mode, proved the presence of a 2-substituted 4-quinolinone moiety. In the COSY spectrum, a spin system corresponding to a (3*Z*,5*E*)-7-hydroxynona-3,5-dien-1-yl moiety was observed. The configuration of the two conjugated double bonds was determined by analysis of the coupling patterns of H-3' and H-4' (H-3': δ 5.44, *dt*, J_{3'-4'} = 10.4 Hz, J_{2'-3'} = 7.7 Hz; H-4': δ 6.03 *br t*, J_{3'-4'} ≈ J_{4'-5'} ≈ 11.1 Hz) and of H-5' and H-6' (H-5': δ 6.45, *ddt*, J_{5'-6'} = 15.2 Hz, J_{4'-5'} = 11.2 Hz, J_{5'-7'} = J_{3'-5'} = 1.2 Hz; H-6': δ 5.62, *dd*, J_{5'-6'} = 15.2 Hz, J_{6'-7'} = 6.3 Hz) as *Z* and *E*, respectively. The hydroxylation of C-7' was established by HSQC correlation of the ¹³C resonance at δ 73.7 with H-7' (δ 3.90), identified by its COSY correlations. Preparative-scale isolation afforded 4 mg of **1**, which showed no optical rotation. Since chiral allylic alcohols and 2,4-diene-1-ols display appreciable specific rotation at the sodium D-line (Airs et al., 1942; Burgess and Jennings, 1991; Takaishi et al., 1991; Tsai et al., 2001), it is concluded that **1** isolated from *H. acutifolium* is a racemate. Compound **1** is a new alkaloid for which the name haplacutine A is suggested.

The material eluted with peak 2 (compound **2a**) was assigned the molecular formula C₁₈H₂₁NO₂ as determined by off-line HRESIMS. Based on the ¹H NMR and COSY spectra acquired in the HPLC–SPE–NMR mode, it contained a 4-quinolinone moiety and a spin system corresponding to a 3-hydroxynona-4,6-dien-1-yl moiety. The configuration of the C-4'–C-5' double bond was established as *E* (H-4': δ 5.70, *br dd*, J_{4'-5'} = 15.2 Hz, J_{3'-4'} = 6.4 Hz; H-5': δ 6.53, *ddt*, J_{4'-5'} = 15.2 Hz, J_{5'-6'} = 11.1 Hz, J_{5'-7'} = J_{3'-5'} = 1.2 Hz) and that of the C-6'–C-7' double bond as *Z* (H-6': δ 5.95, *m*; H-7': δ 5.44, *dt*, J_{6'-7'} = 10.8 Hz, J_{7'-8'} = 7.5 Hz). When the HPLC–PDA–MS–SPE–NMR analysis was performed with the corresponding peak of an alkaloid-enriched fraction of the extract (fraction III obtained from crude extract B, see Section 4), **2a** was accompanied by a closely related analogue **2b** (approximately 60% relative to the major constituent **2a** as determined from the integrals of the ¹H NMR resonances, see Supplementary Fig. 3). The configuration of both double bonds in the minor constituent was *E* (H-4': δ 5.61, *dd*,

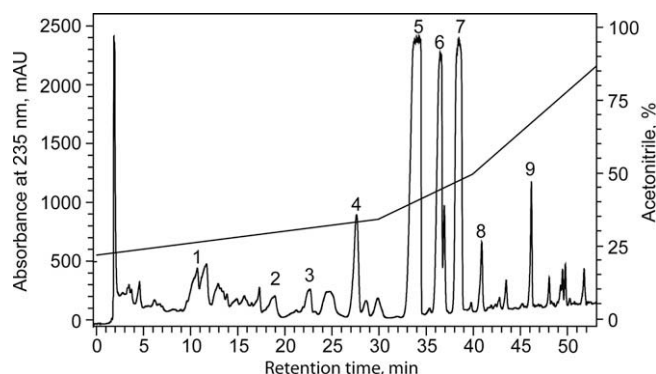


Fig. 1. HPLC profile of raw extract of aerial parts of *Haplophyllum acutifolium* (Phenomenex Luna 2 C18 column, 150 × 4.6 mm i.d., 3 μm, flow rate 0.8 ml/min, MeCN gradient in H₂O + 0.1% HCOOH as shown). The constituents eluted as peaks labeled 1–9 were identified by HPLC–PDA–MS–SPE–NMR experiments.

$J_{4'-5'} = 15.3$ Hz, $J_{3'-4'} = 6.6$ Hz; H-5': δ 6.20, *dd*, $J_{4'-5'} = 15.3$ Hz, $J_{5'-6'} = 10.6$ Hz; H-6': δ 6.05, *dd*, $J_{6'-7'} = 15.3$ Hz, $J_{5'-6'} = 10.6$ Hz; H-7': δ 5.75, *dt*, $J_{6'-7'} = 15.3$ Hz, $J_{7'-8'} = 6.6$ Hz). Thus, **2a** and **2b** are stereoisomers, and they are both new compounds for which the names haplacutine B and haplacutine C, respectively, are suggested. In the crude extract A, **2b** was undetectable, and therefore **2b** co-eluting with **2a** in the alkaloid-enriched fraction III may be an artefact formed by acid-catalyzed, free-radical or photochemical isomerization of **2a** or by an allylic-type rearrangement of **1**.

The constituent eluted as peak 3 (compound **3**) was assigned the molecular formula $C_{18}H_{19}NO_2$ (off-line HRESIFTMS). In addition to signals from a 4-quinolinone moiety, 1H NMR and COSY spectra showed signals corresponding to an aliphatic A_2X_2 spin system and a hexa-1,3-dien-1-yl moiety. The downfield shift of the signals of H-2' and H-5' (δ 3.10 and 7.57, respectively) confirmed that the two moieties are connected by the C-3' carbonyl group. Analysis of the coupling pattern of H-4' to H-7' (see Section 4.9) established the *E*- and *Z*-configuration of the C-4'-C-5' and C-6'-C-7' double bonds, respectively. Compound **3** is a new alkaloid for which the name haplacutine D is suggested.

The constituent eluted as peak 4 (compound **4**) was assigned the molecular formula $C_{16}H_{19}NO$ (HRESIFTMS), and the 1H NMR spectrum showed signals corresponding to a 4-quinolinone moiety and a hept-4-en-1-yl moiety. The 1H NMR data were in agreement with those reported for acutine, previously isolated from *H. acutifolium* (Razzakova et al., 1975). Acutine was initially formulated as 2-(hept-4-enyl)quinolin-4(1*H*)-one (Gulyamova et al., 1974) without any information about the configuration of the double bond, but the coupling patterns observed for H-4' and H-5' in the present investigation (H-4': δ 5.38, *dt*, $J_{4'-5'} = 10.8$ Hz, $J_{3'-4'} = 7.1$ Hz, $J_{4'-6'} = 1.4$ Hz; H-5': δ 5.44, *dt*, $J_{4'-5'} = 10.8$ Hz, $J_{5'-6'} = 7.1$ Hz, $J_{3'-5'} = 1.2$ Hz) proved that compound **4**, and thus acutine, is 2-[(4*Z*)-hept-4-en-1-yl]quinolin-4(1*H*)-one.

Haplamine (**5**) was identified as the material eluted with peak 5. Although the coupling pattern observed for the three hydrogens of the aromatic ring system indicated the presence of a 1,2,4-trisubstituted benzene, the site of methoxylation (i.e., C-6 or C-7) could not be unambiguously assigned by comparison of the observed chemical shift values with those reported for haplamine (syn. 6-methoxyflindersine) (Campbell et al., 1990; Cantrell et al., 2005) and 7-methoxyflindersine (Brader et al., 1996). However, correlations found in a NOESY spectrum acquired in the HPLC-SPE-NMR mode (see Supplementary Fig. 1) proved methoxylation at C-6.

The constituent eluted as peak 6 (compound **6**) was assigned the molecular formula $C_{18}H_{21}NO$ (HRESIFTMS). This is in agreement with the presence of a 4-quinolinone moiety and a nona-3,6-dien-1-yl moiety, as confirmed by connectivities observed in a COSY spectrum. The *Z*-configuration of the C-6'-C-7' double bond was identified based on 1H NMR coupling patterns (H-6': δ 5.20, *dt*, $J_{6'-7'} = 10.7$ Hz, $J_{5'-6'} = 7.3$ Hz, $J_{6'-8'} = 1.6$ Hz; H-7': δ 5.31, *dt*, $J_{6'-7'} = 10.7$ Hz, $J_{7'-8'} = 7.2$ Hz, $J_{5'-7'} = 1.6$ Hz), whereas the configuration of the C-3'-C-4' double bond was not immediately clear because H-3' and H-4' appeared as a multiplet (δ 5.42). However, the simulated coupling pattern of H-3' and H-4' using $J_{3'-4'}$ coupling constant corresponding to the *Z*-configuration was practically identical to the experimentally observed coupling pattern (simulation parameters: $\delta_{H-3'} = 5.43$, $\delta_{H-4'} = 5.41$, $J_{3'-4'} = 10.7$ Hz, $J_{4'-5'} = 7.1$ Hz, $J_{2'-3'} = 6.8$ Hz, $J_{2'-4'} = J_{3'-5'} = 1.1$ Hz), whereas simulated spectrum using an *E* coupling constant was distinctly different (data not shown; cf. however structure of **8** and Fig. 4). In addition, the observed chemical shifts of C-2' and C-5' (δ 27.2 and δ 26.0, respectively, obtained from a HSQC spectrum), were in agreement with the *Z*-configuration of the C-3'-C-4' double bond, whereas chemical shift values above δ 30 would be expected for the *E*-configuration, as shown by chemical shift predictions and literature data (Ivanciuc et al., 1997). Thus, compound **6** is formulated as 2-[(3*Z*,6*Z*)-nona-3,6-dien-1-yl]quinolin-4(1*H*)-one. Full assignment of all 1H and ^{13}C NMR resonances was possible based on 1H , COSY, HSQC, and HMBC experiments obtained in the HPLC-SPE-NMR mode. In order to illustrate the use of HPLC-SPE-NMR methodology, the HSQC and HMBC spectra used for assignment of ^{13}C resonances of **6** are shown in Fig. 3. A compound with a similar structure has previously been isolated from *Vepris ampody* (Kan-Fan et al., 1970). However, the authors were only able to identify the compound as either 2-(nona-3,6-dien-1-yl)quinolin-4(1*H*)-one or 2-(nona-2,6-dien-1-yl)quinolin-4(1*H*)-one, and without any information about configuration of the double bonds. Later, Razzakova et al. (1986) isolated a constituent named 2-nonadie-

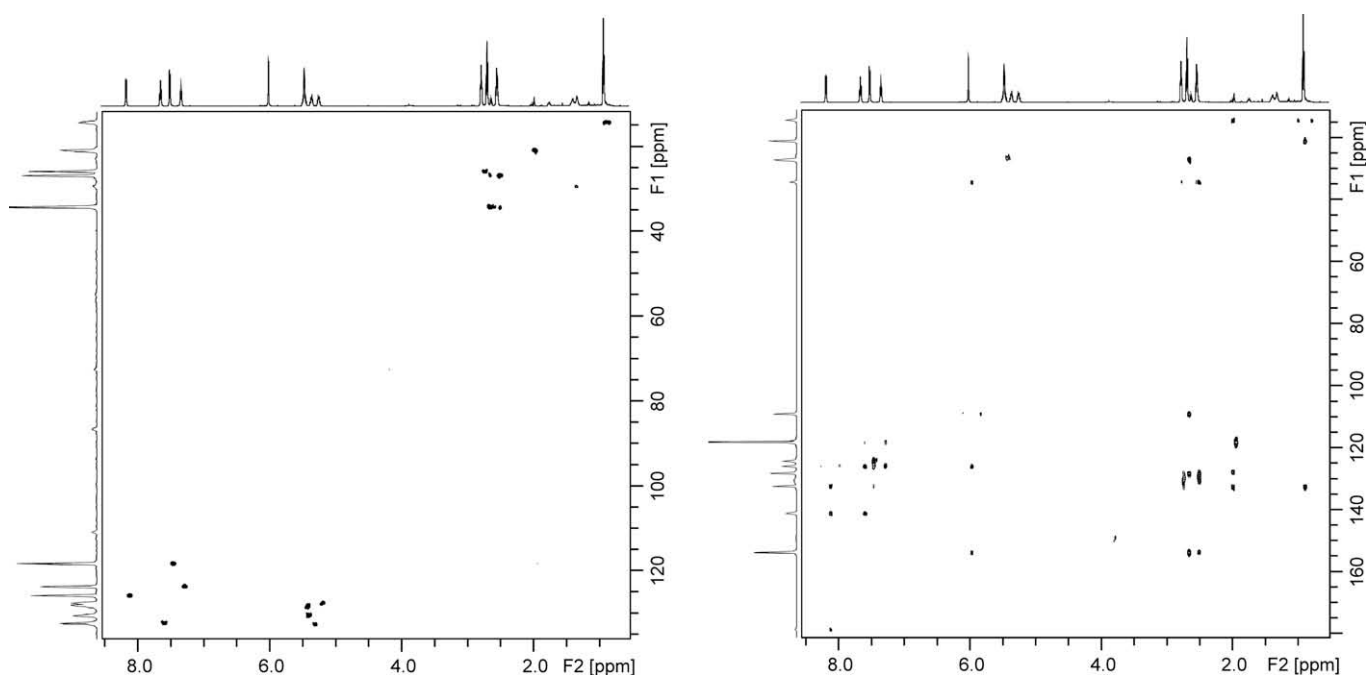


Fig. 3. Gradient-selected HSQC spectrum (left) and HMBC spectrum (right) of compound **6**, allowing full assignment of all ^{13}C resonances in HPLC-SPE-NMR mode.

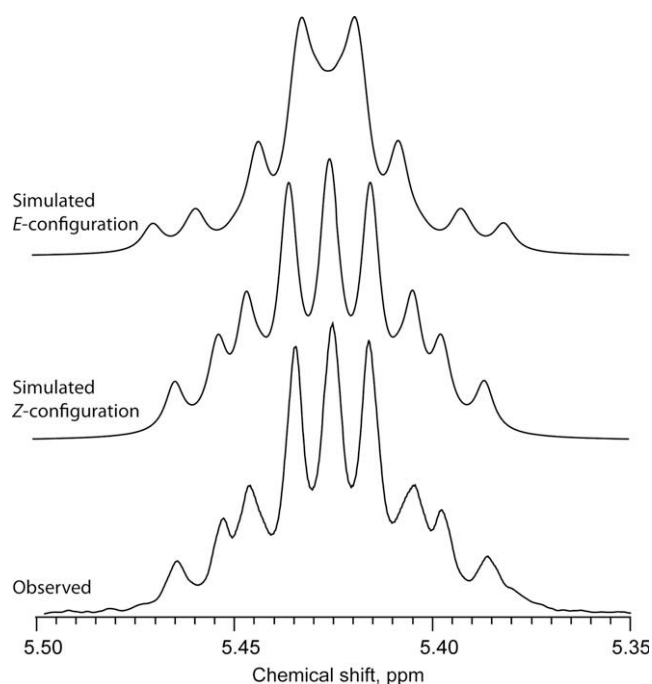


Fig. 4. Coupling pattern for H-3' and H-4' of **8**. Bottom: observed coupling pattern. Middle: simulated coupling pattern for Z-configuration (simulation parameters: $\delta_{3'}$ 5.40, $\delta_{4'}$ 5.44, $J_{3'-4'} = 10.8$ Hz, $J_{3'-2'} = J_{4'-5'} = 10.8$ Hz, $J_{3'-5'} = J_{4'-2'} = 0.7$ Hz). Top: simulated coupling pattern for E-configuration (simulation parameters: as above except $J_{3'-4'} = 15.5$ Hz).

nyl-4-quinoline, but without any information about the position and configuration of the two double bonds. Compound **6** is therefore considered a new compound, for which the name haplacutine E is proposed.

The constituent eluted as peak 7 was identified as (+)-eudesmin (**7**) on the basis of ^1H NMR spectroscopic data recorded in HPLC–SPE–NMR mode (see Supplementary Fig. 2), as well as the identity of ^1H and ^{13}C NMR data and optical rotation data (see Supplementary material) of isolated material with those previously reported (Iida et al., 1982; Miyazawa et al., 1995; Roy et al., 2002). Eudesmine has previously been isolated from *Haplophyllum* species (Razakova et al., 1972).

The constituent eluted as peak 8 (compound **8**) was assigned the molecular formula $\text{C}_{18}\text{H}_{23}\text{NO}$ (HRESIFTMS), and the presence of a 4-quinolinone moiety and a non-3-en-1-yl side chain was revealed by correlations in a COSY spectrum. The Z-configuration was proved by comparing the experimental coupling pattern of the double bond hydrogens with simulated spectra (Fig. 4). This, and the chemical shift values of C-2' and C-5' (δ 27.4 and δ 27.7, respectively), established the structure of **8** as 2-[(3Z)-non-3-en-1-yl]quinolin-4(1H)-one, for which the name haplacutine F is proposed.

2-Nonylquinolin-4(1H)-one (**9**) was identified as the compound eluted as peak 9 based on HRESIFTMS data and comparison of ^1H NMR spectra with those reported in literature (Kostova et al., 1999).

NMR and HRMS data for new compounds are reported in Section 4, whereas retention times, MS data, and NMR data for known compounds are shown in Supplementary Table 1. The spectra of **5**, **7** and **9** obtained in the hyphenation mode are shown in Supplementary Fig. 2. No useful NMR data could be obtained from the remaining peaks. Preparative-scale fractionation of extract B of *H. acutifolium*, comprising a partitioning between chloroform and dilute acid, yielded compounds **1** and **4–7** in small amounts. However, since the *H. acutifolium* alkaloids are tautomeric amides, this standard procedure for alkaloid isolation was not effective, as highlighted by isolation of **6** from the chloroform fraction as well as from the acidic fraction (fractions I and III, respectively,

see Section 4). It was not possible to isolate compound **2–3**, **8**, and **9** in satisfactory amounts and purity. The antiplasmodial activity of compounds **4** and **6** was assessed in an *in vitro* antiplasmodial toxicity assay. Compound **4** and **6** showed IC_{50} values of 2.17 ± 0.22 μM and 3.79 ± 0.24 μM , respectively, whereas the corresponding value for chloroquine diphosphate was 32.8 ± 0.24 nM. Since 4-quinolone alkaloids generally exhibit numerous toxicological activities in the low micromolar range as described in the Introduction, the observed antiplasmodial activity presumably represents a general toxic effect.

3. Conclusion

In conclusion, this study demonstrated the fast and dependable dereplication of alkaloid constituents of crude extract of *H. acutifolium* by HPLC–PDA–MS–SPE–NMR. On-line analysis of nine peaks led to identification of six new and two known 4-quinolinone alkaloids as well as a hemiterpenoid 2-quinolinone alkaloid and a lignan. In agreement with earlier investigations of tautomeric equilibria between hydroxyquinolines and quinolinones in solution (de la Cruz et al., 1992; Mphahlele and El-Nahas, 2004; Tokay and Ögretir, 2002), the alkaloids are formulated as quinolinones. The finding of six new 2-alkylquinolinones in extract of aerial parts of *H. acutifolium* shows that reinvestigation of plants with new and more sensitive technology can add considerable new phytochemical information to already well-investigated species. In continuation of the HPLC–PDA–MS–SPE–NMR investigation, two of the alkaloids were isolated and were shown to inhibit growth of malaria parasites in the low micromolar range. On the basis of this finding, the observed antiplasmodial activity of crude extract of *H. acutifolium* is concluded to represent combined effect of quinolinone alkaloids present in this plant.

4. Experimental

4.1. General

Optical rotations were measured using a Perkin–Elmer 241 polarimeter. NMR spectra of isolated compounds were recorded on a Bruker Avance 400 spectrometer (proton frequency 400.13 MHz) at 25 °C, using TMS as internal standard. HRMS measurements were performed using a Bruker APEX Qe Fourier-transform mass spectrometer equipped with a 9.4 T superconducting cryomagnet and an external ESI source (Apollo II source). The spectra were externally calibrated with arginine cluster in positive and negative ion mode. The samples were dissolved in MeOH, further diluted in 50% MeOH with 0.2% HCOOH, and introduced into the ion source using a syringe pump with a flow of 2 $\mu\text{l}/\text{min}$. Preparative-scale HPLC separations were performed with a 250×16 mm Lichrosorb RP18 (5 μm) column at a flow rate of 8 ml/min, using a chromatograph consisting of a Gynkotec P 580 pump, a Rheodyne 7725 injector, and a Shimadzu SPD-10AV spectrophotometric detector operating at 254 nm, or with a 250×21.2 mm Luna 2 C18 (5 μm) column at a flow rate of 10 ml/min, using a Waters 590 pump, a Rheodyne 7125 injector, and a Lambda-Max Model 481 LC UV detector operating at 235 nm. Spin coupling pattern simulations were performed with gNMR ver. 4.1.2 software (Adept Scientific, Herts, UK). Chemical shift predictions were performed with ChemNMR software as implemented in ChemBioDraw Ultra ver. 11.0 (CambridgeSoft, Cambridge, UK).

4.2. Plant material

Aerial parts of *H. acutifolium* (DC.) G. Don [syn. *Haplophyllum perforatum* (M. Bieb.) Kar. & Kir., *H. flexuosum* Boiss., *H. sieversii*

Fisch., *Ruta perforata* M. Bieb.] (Rutaceae) were collected on August 20, 2004, at a location 40 km west of Mashhad, Iran (altitude 1650 m) and identified by Mr. Ali Ahi. The plant material was dried immediately after collection and powdered before extraction. Voucher specimen (DFHJJ56) was deposited in Herbarium C (Botanical Museum, University of Copenhagen, Copenhagen, Denmark).

4.3. Extraction and sample preparation

Extract A was prepared by macerating 117 g of the plant material with 3×0.5 l of EtOAc, and the combined extracts were evaporated to dryness under reduced pressure to give 2.27 g of residue. A small amount (7.5 mg) of extract A was used for HPLC–PDA–MS–SPE–NMR analysis by making solutions of 28 mg/ml in H₂O–MeCN 2:7, and the rest was used for prep. HPLC. Extract B was prepared by extraction of 500 g of powdered material with 4×1 l of EtOAc and evaporation.

4.4. HPLC–PDA–MS–SPE–NMR experiments

The HPLC–PDA–MS–SPE–NMR instrument consisted of an Agilent 1100 HPLC system (degasser, quaternary solvent delivery pump, autosampler, PDA detector), a Knauer K100 Wellchrom pump for post-column H₂O delivery, a Spark Prospekt 2 solid-phase extraction device, a Bruker EsquireLC mass spectrometer equipped with electrospray ionization (ESI) interface, and a Bruker Avance 600 NMR spectrometer equipped with a 30- μ l inverse ¹H{¹³C} flow-probe operating at 25 °C. HPLC separations were performed at 40 °C on a 150 \times 4.6 mm i.d., 3 μ m particle size, Phenomenex Luna 2 C18 column, using mixtures of H₂O–MeCN 95:5 + 0.1% HCOOH (eluent A) and H₂O–MeCN 5:95 + 0.1% HCOOH (eluent B). The eluate from the column was split with an Accurate splitter, directing 5% of the flow to the mass spectrometer and 95% to the PDA detector. MS spectra were acquired in positive-ion mode, using drying temperature of 350 °C, nebulizer pressure of 15 psi, and drying gas flow of 15 ml/min. The HPLC solvent flow (0.8 ml/min) from the PDA detector was diluted with H₂O (2 ml/min) and directed to the SPE unit. GP phase [general-purpose poly(divinylbenzene)-based resin] SPE cartridges, 10 \times 2 mm i.d. from Spark Holland, were used for trapping of selected compounds based on UV absorption levels at 235 nm. Four cumulative trapings of analytes from separation of 1.4 mg of the raw extract per injection were performed using the following linear gradient elution profile: 0 min, 19% B; 20 min, 32.2% B; 30 min, 50% B; 50 min, 100% B; 60 min, 100% B; 62 min, 19% B, and 8 min conditioning with 19% B. Two extended separations, allowing separation of up to 2.8 mg extract per injection, were performed with four or six adsorptions using the following linear gradient elution profile: 0 min, 19% B; 30 min, 32.2% B; 40 min, 50% B; 60 min, 100% B; 70 min, 100% B; 72 min, 19% B, and 8 min conditioning with 19% B. Prior to use, the SPE cartridges were conditioned with 500 μ l of MeCN at 6 ml/min and flushed with 500 μ l H₂O at 1 ml/min. After trapping, the cartridges were dried with a stream of dry N₂ gas for 45 min and the analytes eluted to the NMR flow cell with CD₃CN. The HPLC–PDA–MS–SPE–NMR experiments were controlled with Bruker HyStar ver. 2.3 software. NMR data acquisition and processing were performed using Bruker XWINNMR ver. 3.1 software. After acquisition of NMR data, the samples were removed from the probe and subjected to HRESIFTMS analysis.

4.5. Preparative-scale isolation

Crude extract B was dissolved in 0.5 l of CHCl₃ and the solution extracted with H₂O acidified to pH 2 with 1 M H₂SO₄ (7 \times 0.5 l) to give fractions I (21 g) and II, respectively. Fraction II was alkalized to pH 11 with concentrated aq. NH₃ and extracted with CHCl₃,

which after concentration yielded 250 mg of fraction III. Fraction I was separated by isocratic prep. HPLC (Lichrosorb RP18, MeCN–H₂O 6:4 + 0.1% TFA), resulting in fractions Ia (20.2 mg), Ib (32.7 mg), and Ic (108.6 mg). These fractions were re-chromatographed to yield 9.6 mg of **6** (Ia: MeCN–H₂O 1:1 + 0.1% TFA), 27.6 mg of **5** (Ib: MeCN–H₂O 4:6 + 0.1% TFA), and 18 mg of **7** (Ic: MeCN–H₂O 6:4 + 0.1% TFA). Fraction III was separated by isocratic prep. HPLC (Lichrosorb RP18, MeCN–H₂O 6:4 + 0.1% TFA), resulting in fractions IIIa–IIIi. Fractions IIIf and IIIi were re-chromatographed using MeCN–H₂O 1:1 + 0.1% TFA to yield 5 mg of **4** and 10 mg of **6**, respectively. Extract A was subjected to targeted isolation of **1** by repeated prep. HPLC (Luna 2 C18, MeCN–H₂O 22:78) to yield 4 mg of **1**.

4.6. Compound **1**

Haplacutine A [2-[(3Z,5E)-7-hydroxynona-3,5-dien-1-yl]quinolin-4(1H)-one]: colourless gum; [α]_D²⁵ 0 (c 0.1, CH₃OH); ¹H NMR spectral data (600 MHz, CD₃OD): δ 0.81 (3H, t, J_{8'-9'} = 7.4, H-9'), 1.37 (1H, dq, J_{8'A-8'B} = 14.2, J_{7'-8'} \approx J_{8'-9'} \approx 7.4, H-8'A), 1.43 (1H, dq, J_{8'A-8'B} = 14.2, J_{7'-8'} \approx J_{8'-9'} \approx 7.4, H-8'B), 2.67 (2H, m, H-2'), 2.81 (2H, m, H-1'), 3.90 (1H, m, H-7'), 5.45 (1H, dt, J_{3'-4'} = 10.6, J_{2'-3'} = 7.7, H-3'), 5.60 (1H, dd, J_{5'-6'} = 15.2, J_{6'-7'} = 6.7, H-6'), 6.06 (1H, t, br, J_{3'-4'} \approx J_{4'-5'} \approx 11.1, H-4'), 6.24 (1H, s, br, H-3), 6.44 (1H, ddt, J_{5'-6'} = 15.2, J_{4'-5'} = 11.2, J_{5'-7'} = J_{3'-5'} = 1.2, H-5'), 7.36 (1H, ddd, J₅₋₆ = 8.0, J₆₋₇ = 7.1, J₆₋₈ = 1.1, H-6), 7.57 (1H, d, br, J₇₋₈ = 8.3, H-8), 7.69 (1H, ddd, J₇₋₈ = 8.3, J₆₋₇ = 7.1, J₅₋₇ = 1.4, H-7), 8.19 (1H, dm, J₅₋₆ = 8.2, H-5); ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 0.81 (3H, t, J_{8'-9'} = 7.4, H-9'), 1.39 (2H, m, H-8'), 2.62 (2H, q, br, J_{1'-2'} \approx J_{2'-3'} \approx 7.4, H-2'), 2.68 (2H, t, br, J_{1'-2'} \approx 7.4, H-1'), 3.90 (1H, q, br, J_{6'-7'} = J_{7'-8'} = 6.3, H-7'), 5.44 (1H, dt, J_{3'-4'} = 10.4, J_{2'-3'} = 7.7, H-3'), 5.62 (1H, dd, J_{5'-6'} = 15.2, J_{6'-7'} = 6.3, H-6'), 5.98 (1H, s, br, H-3), 6.03 (1H, t, br, H-4'), 6.45 (1H, ddt, J_{5'-6'} = 15.2, J_{4'-5'} = 11.2, J_{5'-7'} = J_{3'-5'} = 1.2, H-5'), 7.29 (1H, ddd, J₅₋₆ = 8.0, J₆₋₇ = 7.1, J₆₋₈ = 1.0, H-6), 7.46 (1H, d, br, J₇₋₈ = 8.3, H-8), 7.60 (1H, ddd, J₇₋₈ = 8.3, J₆₋₇ = 7.1, J₅₋₇ = 1.4, H-7), 8.11 (1H, dd, J₅₋₆ = 8.0, J₅₋₇ = 1.4, H-5); ¹³C NMR spectral data (150 MHz, CD₃CN, from HSQC spectrum recorded in hyphenation mode): δ 10.1 (C-9'), 27.4 (C-2'), 30.8 (C-8'), 34.1 (C-1'), 73.7 (C-7'), 109.2 (C-3), 118.3 (C-8), 123.9 (C-6), 125.0 (C-5'), 126.0 (C-5), 129.6 (C-3'), 130.4 (C-4'), 132.5 (C-7), 138.7 (C-6'); HRESIFTMS m/z 284.16467 [MH]⁺ (calc. for [C₁₈H₂₂NO₂]⁺, 284.16451).

4.7. Compound **2a**

Haplacutine B [2-[(4E,6Z)-3-hydroxynona-4,6-dien-1-yl]quinolin-4(1H)-one]: ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 0.98 (3H, t, J_{8'-9'} = 7.5, H-9'), 1.87 (2H, m, H-2'), 2.09 (2H, m, H-8'), 2.66 (2H, m, H-1'), 3.11 (1H, d, br, J_{3'-3'-OH} \approx 4.1, 3'-OH), 4.18 (1H, s, br, H-3'), 5.44 (1H, dt, J₆₋₇ = 10.8, J_{7'-8'} = 7.5, H-7'), 5.70 (1H, dd, br, J_{4'-5'} = 15.2, J_{3'-4'} = 6.4, H-4'), 5.95 (1H, m, H-6'), 5.98 (1H, s, br, H-3), 6.53 (1H, ddt, J_{4'-5'} = 15.2, J_{5'-6'} = 11.1, J_{5'-7'} = J_{3'-5'} = 1.2, H-5'), 7.28 (1H, ddd, J₅₋₆ = 8.0, J₆₋₇ = 7.0, J₆₋₈ = 1.0, H-6), 7.43 (1H, d, br, J₇₋₈ = 8.4, H-8), 7.59 (1H, ddd, J₇₋₈ = 8.4, J₆₋₇ = 7.0, J₅₋₇ = 1.5, H-7), 8.12 (1H, d, br, J₅₋₆ = 8.0, H-5); HRESIFTMS m/z 284.16460 [MH]⁺ (calc. for [C₁₈H₂₂NO₂]⁺, 284.16451).

4.8. Compound **2b**

Haplacutine C [2-[(4E,6E)-3-hydroxynona-4,6-dien-1-yl]quinolin-4(1H)-one]: ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 0.99 (3H, t, J_{8'-9'} = 7.5, H-9'), 1.87 (2H, m, H-2'), 2.19 (2H, pd, J_{8'-9'} = J_{7'-8'} = 7.5, J_{6'-8'} = 1.5, H-8'), 2.70 (2H, m, H-1'), 3.06 (1H, d, br, J_{3'-3'-OH} \approx 4.1, 3'-OH), 4.13 (1H, s, br, H-3'), 5.61 (1H, dd, J_{4'-5'} = 15.3, J_{3'-4'} = 6.6, H-4'), 5.75 (1H, dt, J_{6'-7'} = 15.3, J_{7'-8'} = 6.6, H-7'), 5.98 (1H, s, br, H-3), 6.05 (1H, dd, J_{6'-7'} = 15.3,

$J_{5'-6'} = 10.6$, H-6'), 6.20 (1H, *dd*, $J_{4'-5'} = 15.3$, $J_{5'-6'} = 10.6$, H-5'), 7.28 (1H, *ddd*, $J_{5-6} = 8.0$, $J_{6-7} = 7.0$, $J_{6-8} = 1.0$, H-6), 7.43 (1H, *d*, *br*, $J_{7-8} = 8.4$, H-8), 7.59 (1H, *ddd*, $J_{7-8} = 8.4$, $J_{6-7} = 7.0$, $J_{5-7} = 1.5$, H-7), 8.12 (1H, *d*, *br*, $J_{5-6} = 8.0$, H-5); HRESIFTMS m/z 284.16460 [MH]⁺ (calc. for [C₁₈H₂₂NO₂]⁺, 284.16451).

4.9. Compound 3

Haplaclutine D [2-[(4E,6Z)-3-oxonona-4,6-dien-1-yl]quinolin-4(1H)-one]: ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 1.02 (3H, *t*, $J_{8'-9'} = 7.5$, H-9'), 2.34 (2H, *pd*, $J_{8'-9'} = J_{7'-8'} = 7.5$, $J_{6'-8'} = 1.4$, H-8'), 2.88 (2H, *t*, $J_{1'-2'} = 7.3$, H-1'), 3.10 (2H, *t*, $J_{1'-2'} = 7.3$, H-2'), 5.96 (1H, *dt*, $J_{6'-7'} = 11.1$, $J_{7'-8'} = 7.5$, H-7'), 5.97 (1H, *s*, *br*, H-3), 6.15 (1H, *t*, *br*, $J_{6'-7'} \approx J_{5'-6'} \approx 11.1$, H-6'), 6.20 (1H, *d*, $J_{4'-5'} = 15.5$, H-4'), 7.29 (1H, *t*, *br*, $J_{5-6} \approx J_{6-7} = 7.6$, H-6), 7.44 (1H, *d*, *br*, $J_{7-8} = 8.2$, H-8), 7.57 (1H, *dd*, $J_{4'-5'} = 15.5$, $J_{5'-6'} = 11.5$, H-5'), 7.61 (1H, *dd*, $J_{7-8} = 8.2$, $J_{6-7} = 7.6$, H-7), 8.11 (1H, *d*, *br*, $J_{5-6} = 8.2$, H-5); HRESIFTMS m/z 282.14885 [MH]⁺ (calc. for [C₁₈H₂₀NO₂]⁺, 282.14886).

4.10. Compound 6

Haplaclutine E [2-[(3Z,6Z)-nona-3,6-dien-1-yl]quinolin-4(1H)-one]: ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 0.89 (3H, *t*, $J_{8'-9'} = 7.5$, H-9'), 1.99 (2H, *d*, *br*, H-8'), 2.50 (2H, *dt*, $J_{2'-3'} = 6.8$, $J_{1'-2'} = 7.5$, H-2'), 2.66 (2H, *t*, $J_{1'-2'} = 7.5$, H-1'), 2.74 (2H, *t*, *br*, $J_{4'-5'} \approx J_{5'-6'} \approx 6.5$, H-5'), 5.20 (1H, *dt*, $J_{6'-7'} = 10.7$, $J_{5'-6'} = 7.3$, $J_{6'-8'} = 1.6$ Hz, H-6'), 5.31 (1H, *dt*, $J_{6'-7'} = 10.7$, $J_{7'-8'} = 7.2$, $J_{5'-7'} = 1.6$, H-7'), 5.42 (2H, *m*, H-3', H-4'), 5.97 (1H, *s*, *br*, H-3), 7.29 (1H, *t*, *br*, $J_{5-6} \approx J_{6-7} \approx 7.7$, H-6), 7.46 (1H, *d*, *br*, $J_{7-8} = 8.3$, H-8), 7.60 (1H, *ddd*, $J_{7-8} = 8.3$, $J_{6-7} = 7.0$, $J_{5-7} = 1.6$, H-7), 8.12 (1H, *dd*, $J_{5-6} = 8.1$, $J_{5-7} = 1.2$, H-5); ¹³C NMR spectral data (150 MHz, CD₃CN): δ 14.4 (C-9'), 21.0 (C-8'), 25.9 (C-5'), 26.9 (C-2'), 34.3 (C-1'), 109.0 (C-3), 118.2 (C-8), 123.8 (C-6), 125.9 (C-5), 126.1 (C-10), 127.8 (C-6'), 128.4 (C-3'), 130.6 (C-4'), 132.3 (C-7), 132.7 (C-7'), 141.2 (C-9), 154.0 (C-2), 178.7 (C-4); HRESIFTMS m/z 268.16965 [MH]⁺ (calc. for [C₁₈H₂₂NO]⁺, 268.16959).

4.11. Compound 8

Haplaclutine F [2-[(3Z)-non-3-en-1-yl]quinolin-4(1H)-one]: ¹H NMR spectral data (600 MHz, CD₃CN, hyphenation mode): δ 0.80 (3H, *t*, $J_{8'-9'} = 7.0$, H-9'), 1.16 (4H, *m*, H-7', H-8'), 1.20 (2H, *m*, H-6'), 1.98 (*m*, H-5'), 2.47 (2H, *q*, *br*, $J_{1'-2'} \approx J_{2'-3'} \approx 7.0$, H-2'), 2.64 (2H, *t*, $J_{1'-2'} = 7.4$, H-1'), 5.42 (2H, *m*, H-3', H-4'), 5.95 (1H, *s*, *br*, H-3), 7.28 (1H, *t*, *br*, $J_{5-6} \approx J_{6-7} \approx 7.5$, H-6), 7.45 (1H, *d*, *br*, $J_{7-8} = 8.3$, H-8), 7.59 (1H, *ddd*, $J_{7-8} = 8.3$, $J_{6-7} = 7.0$, $J_{5-7} = 1.4$, H-7), 8.11 (1H, *d*, *br*, $J_{5-6} = 8.0$, H-5), 9.47 (1H, *s*, *br*, H-1); ¹³C NMR (150 MHz, CD₃CN): δ 23.3 (C-8'), 27.4 (C-2'), 27.7 (C-5'), 29.8 (C-6'), 31.9 (C-7'), 34.5 (C-1'), 109.1 (C-3), 118.2 (C-8), 124.0 (C-6), 126.1 (C-5), 132.4 (C-7); HRESIFTMS m/z 270.18521 [MH]⁺ (calc. for [C₁₈H₂₄NO]⁺, 270.18524).

4.12. Antiplasmodial assay

Chloroquine-sensitive *P. falciparum* 3D7 strain was cultured *in vitro* using a modified method of Trager and Jensen (1976), and quantitative assessment of antiplasmodial activity *in vitro* was performed as described in details elsewhere (Sairafianpour et al., 2001; Ziegler et al., 2002).

Acknowledgments

We thank Ms. Birgitte Simonsen and Ms. Dorte Brix (Department of Medicinal Chemistry, University of Copenhagen) for tech-

nical assistance. Postdoctoral fellowship to MS from Apotekerfonden af 1991 (Copenhagen) is gratefully acknowledged.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.phytochem.2009.05.004.

References

- Airs, R.S., Balfe, M.P., Keynon, J., 1942. Reactions of (+)- and (−)-γ-methyl-α-ethylallyl alcohols and their derivatives. *J. Chem. Soc.* 1, 8–26.
- Albert, K., 2002. On-line NMR and Related Techniques. Chichester, Wiley.
- Bieri, S., Varesio, E., Veuthey, J.L., Munoz, O., Tseng, L.H., Braumann, U., Spraul, M., Christen, P., 2006. Identification of isomeric tropane alkaloids from *Schizanthus grahamii* by HPLC–NMR with loop storage and HPLC–UV–MS/SPE–NMR using a cryogenic flow probe. *Phytochem. Anal.* 17, 78–86.
- Brader, G., Bacher, M., Greger, H., Hofer, O., 1996. Pyranoquinolones and acridones from *Vepris bilocularis*. *Phytochemistry* 42, 881–884.
- Burgess, K., Jennings, L.D., 1991. Enantioselective esterification of unsaturated alcohols mediated by a lipase prepared from *Pseudomonas* sp. *J. Am. Chem. Soc.* 113, 6129–6139.
- Campbell, W.E., Davidowitz, B., Jackson, G.E., 1990. Quinolone alkaloids from an *Agathosma* species. *Phytochemistry* 29, 1303–1306.
- Cantrell, C.L., Schrader, K.K., Mamonov, L.K., Sitpaeva, G.T., Kustova, T.S., Dunbar, C., Wedge, D.E., 2005. Isolation and identification of antifungal and antialgal alkaloids from *Haplophyllum sieversii*. *J. Agric. Food Chem.* 53, 7741–7748.
- Cheng, M.J., Lee, K.H., Tsai, I.L., Chen, I.S., 2005. Two new sesquiterpenoids and anti-HIV principles from the root bark of *Zanthoxylum ailanthoides*. *Bioorg. Med. Chem.* 13, 5915–5920.
- Clarkson, C., Staerk, D., Hansen, S.H., Jaroszewski, J.W., 2005. Hyphenation of solid-phase extraction with liquid chromatography and nuclear magnetic resonance. Application of HPLC–DAD–SPE–NMR to identification of constituents of *Kanahia laniflora*. *Anal. Chem.* 77, 3547–3553.
- Clarkson, C., Staerk, D., Hansen, S.H., Smith, P.J., Jaroszewski, J.W., 2006a. Discovering new natural products directly from crude extracts by HPLC–SPE–NMR: chinane diterpenes in *Harpagophytum procumbens*. *J. Nat. Prod.* 69, 527–530.
- Clarkson, C., Staerk, D., Hansen, S.H., Smith, P.J., Jaroszewski, J.W., 2006b. Identification of major and minor constituents of *Harpagophytum procumbens* (Devil's Claw) using HPLC–SPE–NMR and HPLC–ESIMS/APCIMS. *J. Nat. Prod.* 69, 1280–1288.
- Clarkson, C., Sibum, M., Mensen, R., Jaroszewski, J.W., 2007. Evaluation of on-line solid-phase extraction parameters for hyphenated, high-performance liquid chromatography–solid-phase extraction–nuclear magnetic resonance applications. *J. Chromatogr. A* 1165, 1–9.
- de la Cruz, A., Elguero, J., Goya, P., Martinez, A., Pfeleiderer, W., 1992. Tautomerism and acidity in 4-quinolone-3-carboxylic acid derivatives. *Tetrahedron* 48, 6135–6150.
- Exarchou, V., Fiamegos, Y.C., van Beek, T.A., Nanos, C., Vervoort, J., 2006. Hyphenated chromatographic techniques for the rapid screening and identification of antioxidants in methanolic extracts of pharmaceutically used plants. *J. Chromatogr. A* 1112, 293–302.
- Gulyamova, D.M., Bessonova, I.A., Yunusov, S.Y., 1974. Alkaloids of *Haplophyllum acutifolium*. *Chem. Nat. Compd.* 7, 836–837.
- Hanawa, F., Fokialakis, N., Skaltsounis, A.L., 2004. Photo-activated DNA binding and antimicrobial activities of furoquinoline and pyranoquinolonealkaloids from Rutaceae. *Planta Med.* 70, 531–535.
- Iida, T., Nakano, M., Ito, K., 1982. Hydroperoxy sesquiterpene and lignan constituents of *Magnolia kobus*. *Phytochemistry* 21, 673–675.
- Ivanciuc, O., Rabine, J.P., Cabrol-Bass, D., Panaye, A., Doucet, J.P., 1997. ¹³C NMR chemical shift prediction of the sp³ carbon atoms in the α position relative to the double bond in acyclic alkenes. *J. Chem. Inf. Comput. Sci.* 37, 587–598.
- Jaroszewski, J.W., 2005a. Hyphenated NMR methods in natural products research, part 1: direct hyphenation. *Planta Med.* 71, 691–700.
- Jaroszewski, J.W., 2005b. Hyphenated NMR methods in natural products research, part 2: HPLC–SPE–NMR and other new trends in NMR hyphenation. *Planta Med.* 71, 795–802.
- Kan-Fan, C., Das, B.C., Boiteau, P., Potier, P., 1970. Alcaloïdes de *Vepris ampody* (Rutacées). *Phytochemistry* 9, 1283–1291.
- Kostova, I., Ivanova, A., Mikhova, B., Klaiher, I., 1999. Alkaloids and coumarins from *Ruta graveolens*. *Monatsh. Chem.* 130, 703–707.
- Lambert, M., Staerk, D., Hansen, S.H., Sairafianpour, M., Jaroszewski, J.W., 2005a. Rapid extract dereplication using HPLC–SPE–NMR: analysis of isoflavonoids from *Smirnowia iranica*. *J. Nat. Prod.* 68, 1500–1509.
- Lambert, M., Staerk, D., Hansen, S.H., Jaroszewski, J.W., 2005b. HPLC–SPE–NMR hyphenation in natural products research: optimization of analysis of *Croton membranaceus* extract. *Magn. Reson. Chem.* 43, 771–775.
- Lambert, M., Wolfender, J.-L., Staerk, D., Christensen, S.B., Hostettmann, K., Jaroszewski, J.W., 2007. Identification of natural products using HPLC–SPE combined with CapNMR. *Anal. Chem.* 79, 727–735.

- Lee, S.S., Lai, Y.-C., Chen, C.-K., Tseng, L.-H., Wang, C.-Y., 2007. Characterization of isoquinoline alkaloids from *Neolitsea sericea* var. *aurata* by HPLC–SPE–NMR. *J. Nat. Prod.* 70, 637–642.
- Michael, J.P., 2003. Quinoline, quinazoline and acridone alkaloids. *Nat. Prod. Rep.* 20, 476–493.
- Michael, J.P., 2008. Quinoline, quinazoline and acridone alkaloids. *Nat. Prod. Rep.* 25, 166–187.
- Miyazawa, M., Kasahara, H., Kameoka, H., 1995. Biotransformation of lignans: metabolism of (+)-eudesmin and (+)-magnolin in *Spodoptera litura*. *Phytochemistry* 39, 1027–1030.
- Mphahlele, M.J., El-Nahas, A.M., 2004. Tautomeric 2-arylquinolin-4(1H)-one derivatives- spectroscopic, X-ray and quantum chemical structural studies. *J. Mol. Struct.* 688, 129–136.
- Nazrullaev, S.S., Bessonova, I.A., Akhmedkhodzhaeva, Kh.S., 2001. Estrogenic activity as a function of chemical structure in *Haplophyllum* quinoline alkaloids. *Chem. Nat. Compd.* 37, 551–555.
- Razzakova, D.M., Bessonova, I.A., Yunusov, S.Y., 1972. Eudesmine – a lignane from *Haplophyllum acutifolium* and *H. perforatum*. *Chem. Nat. Compd.* 8, 646–647.
- Razzakova, D.M., Bessonova, I.A., Yunusov, S.Y., 1975. Alkaloids of *Haplophyllum acutifolium*. *Chem. Nat. Compd.* 9, 199–202.
- Razzakova, D.M., Bessonova, I.A., Yunusov, S.Y., 1986. Alkaloids of *Haplophyllum acutifolium*. *Chem. Nat. Compd.* 22, 363–364.
- Roy, S.C., Rana, K.K., Guin, C., 2002. Short and stereoselective total synthesis of furano lignans (±)-dihydrosesamin, (±)-lariciresinol dimethyl ether, (±)-acuminatin methyl ether, (±)-sanskodiol methyl ether, (±)-lariciresinol, (±)-acuminatin, and (±)-lariciresinol monomethyl ether and furofuran lignans (±)-sesamin, (±)-eudesmin, (±)-piperitol methyl ether, (±)-pinioresinol, (±)-piperitol, and (±)-pinioresinol monomethyl ether by radical cyclization of epoxides using a transition-metal radical source. *J. Org. Chem.* 67, 3242–3248.
- Sairafianpour, M., Christensen, J., Staerk, D., Budnik, B.A., Kharazmi, A., Bagherzadeh, K., Jaroszewski, J.W., 2001. Leishmanicidal, antiplasmodial, and cytotoxic activity of novel diterpenoid 1, 2-quinones from *Perovskia abrotanoides*: new source of tanshinones. *J. Nat. Prod.* 64, 1398–1403.
- Sprogø, K., Staerk, D., Jäger, A.K., Adersen, A., Hansen, S.H., Witt, M., Landbo, A.-K.R., Meyer, A.S., Jaroszewski, J.W., 2007. Targeted natural product isolation guided by HPLC–SPE–NMR: constituents of *Hubertia* species. *J. Nat. Prod.* 70, 1472–1477.
- Staerk, D., Lambert, M., Jaroszewski, J.W., 2006. HPLC–NMR techniques for plant extract analysis. In: Kayser, O., Quax, W. (Eds.), *Medicinal Plant Biotechnology*, Vol. 1. Wiley-VCH, Weinheim, pp. 29–48.
- Sturm, S., Seger, C., 2007. HPLC–SPE–NMR – A novel hyphenation technique. *LCGC Europe*, 587–597.
- Sturm, S., Seger, C., Godejohann, M., Spraul, M., Stuppner, H., 2007. Conventional sample enrichment strategies combined with high-performance liquid chromatography–solid phase extraction–nuclear magnetic resonance analysis allows analyte identification from a single minuscule *Corydalis solida* plant tuber. *J. Chromatogr. A* 1163, 138–144.
- Takaishi, Y., Okuyama, T., Nakano, K., Murakami, K., Tomimatsu, T., Yamahara, J., 1991. Polyolefinic compounds from *Cirsium nipponicum* roots. *Phytochemistry* 30, 1539–1542.
- Tatsis, E.C., Boeren, S., Exarchou, V., Troganis, A.N., Vervoort, J., Gerothanassis, I.P., 2007. Identification of the major constituents of *Hypericum perforatum* by LC/SPE/NMR and/or LC/MS. *Phytochemistry* 68, 383–393.
- Tokay, N., Ögretir, C., 2002. Quantum chemical studies on tautomerism of 2-, 3- or 4-hydroxyquinoline derivatives along with their thio and azo analogs. *J. Mol. Struct.* 594, 185–197.
- Trager, W., Jensen, J.B., 1976. Human malaria parasite in continuous culture. *Science* 193, 673–675.
- Tsai, I.-L., Lin, W.-Y., Huang, M.-W., Chen, T.-L., Chen, I.-S., 2001. *N*-Isobutylamides and butyrolactone from the fruits of *Zanthoxylum integrifolium*. *Helv. Chim. Acta* 84, 830–883.
- Ziegler, H.L., Staerk, D., Christensen, J., Hviid, L., Hägerstrand, H., Jaroszewski, J.W., 2002. *In vitro* *Plasmodium falciparum* drug sensitivity assay: inhibition of parasite growth by incorporation of stomatocytogenic amphiphiles into the erythrocyte membrane. *Antimicrob. Agents Chemother.* 46, 1441–1446.